



SensoLyte® 490 MMP-7 Assay Kit *Fluorimetric*

Catalog #	71132
Unit Size	1 kit
Kit Size	500 assays (96-well) or 1250 assays (384-well)

This kit is optimized to detect MMP-7 activity using an NBD/DMC FRET peptide as substrate, which can be monitored at Ex/Em=370 nm/460 nm upon proteolytic cleavage. Sufficient materials are provided to perform 500 assays in a 96-well format or 1250 assays in a 384-well format. The kit has the following features:

- **Convenient Format:** All essential assay components are included.
- **Optimized Performance:** Optimal conditions for the detection of MMP-7 activity.
- **Enhanced Value:** Less expensive than the sum of individual components.
- **High Speed:** Minimal hands-on time.
- **Assured Reliability:** Detailed protocol and references are provided.

USA and Canada Ordering Information

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Tel: 408-452-5055
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International Ordering Information

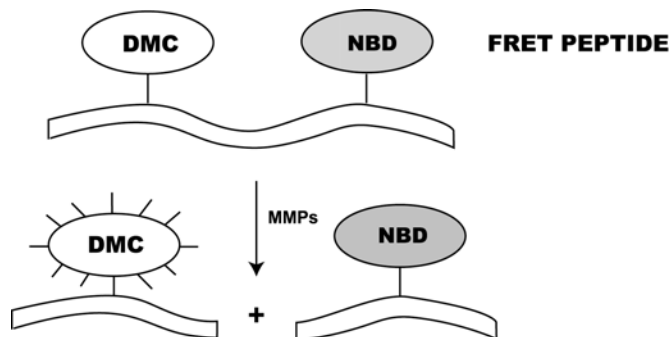
A list of international distributors is available at www.anaspec.com.

INTRODUCTION

Matrix metalloproteinases (MMPs) belong to a family of secreted or membrane-associated zinc endopeptidases capable of digesting extracellular matrix components.^{1,2} The importance of MMPs in tumor development and invasion as well as other diseases is well known. MMP-7,³ (matrilysin, PUMP) is proposed as a potential anti-cancer drug target.

The SensoLyte® 490 MMP-7 Assay Kit provides a convenient assay for high throughput screening of MMP-7 inducers and inhibitors. It detects MMP-7 activity in a variety of biological samples using a DMC/NBD fluorescence resonance energy transfer (FRET)⁴ peptide. In the intact FRET peptide, the fluorescence of DMC is quenched by NBD. Upon cleavage into two separate fragments by MMP-7 (**Scheme 1**), the fluorescence of DMC is recovered, and can be monitored at excitation/emission wavelengths = 370 nm/460 nm.

The assays are performed in a convenient 96-well or 384-well microplate format.



Scheme 1. Proteolytic cleavage of DMC/NBD FRET peptide by MMPs.

KIT COMPONENTS, STORAGE AND HANDLING

Note: Store all kit Components at -20 °C, and keep Components A and B from light. If used frequently, Components D and E can be stored at 4 °C for convenience.

- Component A:** MMP-7 substrate (270 µL)
DMC/NBD FRET Peptide
Ex/Em=370 nm/460 nm upon cleavage
- Component B:** DMC, fluorescence reference standard (1 mM DMSO solution, 10 µL)
Ex/Em=370 nm/460 nm
- Component C:** APMA, 4-aminophenylmercuric acetate (1 M, 100 µL)
Caution: Organic mercury. Handle with care! Do not dump into sink!
- Component D:** Assay buffer (60 mL)
- Component E:** Stop solution (30 mL)

OTHER MATERIALS REQUIRED (BUT NOT PROVIDED)

Recombinant MMP-7: AnaSpec Cat#72007.

96-well or 384-well microplate: Black microplate provides better signal/noise value.

Fluorescence microplate reader: Capable of detecting emission at 460 nm with excitation at 370 nm.

PROTOCOL

Note 1: For fluorometer calibration, please refer to Appendix II (recommended for the first time users).

Note 2: Please use protocol A or B based on your needs.

Protocol A. Screening protease inhibitors using purified or recombinant MMP-7.

1. Activate pro-MMP-7

- Incubate pro-MMP-7 with 1 mM APMA (diluted component C) for 1 h at 37°C. Activate pro-MMP-7 immediately before the experiment.

Note 1: Keep activated enzyme on ice. Avoid vigorously vortexing the enzyme. Prolonged storage of activated enzyme will further de-activate the enzyme.

Note 2: APMA can be diluted with assay buffer (component D). APMA belongs to organic mercury. Handle with care! Dispose it according to appropriate regulations.

Note 3: Activation of zymogen by APMA at higher protein concentration. After activation, the enzyme may be further diluted.

2. Prepare working solutions

Note: Warm all kit components until thawed to room temperature before starting the experiments.

- **MMP-7 substrate solution:** Dilute MMP-7 substrate (component A) 1: 100 in assay buffer (component D).

Table 1. MMP-7 substrate solution for one 96-well plate (100 assays).

Components	Volume
MMP-7 substrate (100X, component A)	50 μ L
Assay buffer (component D)	4.95 mL
Total volume	5 mL

- **MMP-7 diluent:** Dilute activated MMP-7 to appropriate concentration in assay buffer (component D).

3. Set up enzymatic reaction.

- Add test compounds and MMP-7 diluent into microplate. The suggested total volume of MMP-7 diluent and test compound is 50 μ L (96 well plate) or 20 μ L (384-well plate).
- Simultaneously set up the following controls.
 - **Positive control** contains MMP-7 diluent without test compound.
 - **Inhibitor control** contains MMP-7 diluent and a known MMP-7 inhibitor.
 - **Vehicle control** contains MMP-7 diluent and vehicle used in delivering test compound (e.g. DMSO).
 - **Test compound control** contains assay buffer and test compound. Some test compounds have strong autofluorescence and may give false results.
 - **Substrate control** contains assay buffer only.

Note: Use assay buffer (Component D) to bring the total volume of all the controls to 50 μ L (96-well plate) or 20 μ L (384-well).

4. Pre-incubation.

- Incubate the plate at the desired temperature for enzymatic reaction (e.g. 25°C or 37°C) for 10-15 min. In the mean time, also incubate MMP-7 substrate solution at the same temperature.

5. Initiate the enzymatic reaction.

- Add 50 μ L of MMP-7 substrate solution into the wells of a 96-well plate. Or add 20 μ L into the wells of a 384-well plate. Mix the reagents completely by shaking the plate gently for 30-60 second.
- Measure fluorescence signal:

For kinetic reading: Immediately start measuring fluorescence intensity at Ex/Em=340±30 nm/490±30 nm continuously and record data every 5 minutes for 30 to 60 minutes.

For end-point reading: Incubate the reaction at room temperature for 30 to 60 minutes. Keep the plate from direct light. (Optional) add 50 µL/well stop solution (component E) to 96-well plate or 20 µL/well to 384-well plate. Mix the reagents. Then measure fluorescence intensity at Ex/Em=340±30 nm/490±30 nm.

- Data analysis: Refer to Appendix I.

Protocol B. Measuring MMP-7 activity in biological samples.

Note: The FRET substrate in this kit can also be cleaved by MMP-1, 12, and 13. If several MMPs are coexisting in your samples and you want to specifically measure MMP-7's activity, then MMP-7 must first be isolated by immuno-affinity purification or other methods before measuring its specific activity using current assay kit.

Prepare MMP-7 containing biological samples.

- Collect synovial fluids or supernatant of cell culture media (e.g. stimulated fibroblast) and centrifuge for 10-15 min at 1,000X g, 4°C. Collect the supernatant and store at -70°C until use.
- Tissues samples should be homogenized in assay buffer (Component D) containing 0.1% Triton X-100, and then centrifuged for 15 min at 10000x g at 4°C. Collect the supernatant and store at -70°C until use.

Note: Triton-X 100 is not provided.

2. Activate pro-MMPs.

- Incubate the MMP containing-samples with APMA (Component C) at the final concentration of 1 mM in the assay buffer (component D) for 1 h at 37°C. Activate MMP right before the experiment.
Note 1: Keep activated enzyme on ice. Avoid vigorously vortexing the enzyme. Prolonged storage will further de-activate the enzyme.
Note 2: APMA can be diluted with assay buffer (component D). APMA belongs to organic mercury. Handle with care! Dispose it according to appropriate regulations.

3. Prepare working solutions.

Note: Warm all kit components until thawed to room temperature before starting the experiments.

- MMP-7 substrate solution: Dilute MMP-7 substrate (component A) 1: 100 in assay buffer (component D).

Table 1. MMP-7 substrate solution for one 96-well plate (100 assays).

Components	Volume
MMP-7 substrate (100X, component A)	50 µL
Assay buffer (component D)	5 mL
Total volume	5 mL

- MMP-7 diluent: If you use purified MMP-7 as positive control, then dilute MMP-7 to an appropriate concentration in assay buffer (Component D).

Note: Pro-MMP 7 needs to be activated by APMA at higher concentration, and then diluted to a working concentration in assay buffer. Please refer to Step 2. Avoid vigorous vortexing of enzyme.

4. Set up the enzymatic reaction.

- Add 50 µL/well MMP-7 containing sample to 96-well plate (20 µL/well to 384-well plate).
- Set up the following control:
 - Substrate control contains assay buffer (50 µL/well for a 96-well plate or 20 µL/well for a 384-well plate).
 - Positive control: contains MMP-7 diluent (50 µL/well for a 96-well plate or 20 µL/well for a 384-well plate).

5. Initiate the enzymatic reaction.

- Add 50 µL/well MMP-7 substrate solution to the sample and control wells of 96-well plate. Or add 20 µL/well to 384-well plate. Mix the reagents by shaking the plate gently for 30 seconds.
- Measure fluorescence signal:

For kinetic reading: Immediately start measuring fluorescence intensity at Ex/Em=370 nm/460 nm continuously and record data every 5 minutes for 30 to 60 minutes.

For end-point reading: Incubate the reaction at room temperature for 30 to 60 minutes. Keep the plate from direct light. (Optional) add 50 μL /well stop solution (component E) to 96-well plate or 20 μL /well to 384-well plate. Mix the reagents. Then measure fluorescence intensity at Ex/Em=370 nm/460 nm.

- Data analysis: Refer to Appendix I.

Appendix I. Data Analysis

- The fluorescence reading from the substrate control well is the background fluorescence. This background reading has to be subtracted from the readings of the other wells. This reading is the relative fluorescence unit (RFU).
- For kinetic reading:
 - Plot data as RFU versus time for each sample. If you want to convert the RFU to the concentration of the product of enzymatic reaction, please refer to [Appendix II](#) for setting up fluorescence reference standard.
 - Determine the range of initial time points during which the reaction is linear. 10-15% conversion appears to be the optimal range.
 - Obtain the initial reaction velocity (V_0) in RFU/min. Determine the slope of the linear portion of the data plot.
 - A variety of data analyses can be done, e.g., determining inhibition %, EC_{50} , IC_{50} , K_m , K_i , etc.
- For endpoint reading:
 - Plot data as RFU versus the concentration of test compounds.
 - A variety of data analyses can be done, e.g., determining inhibition %, EC_{50} , IC_{50} , etc.

Appendix II: Fluorometer calibration.

- **DMC fluorescence reference standard:** dilute 1 mM DMC (component B) to 5 μM in deionized water. Do 1:2 serial dilutions to get concentrations of 2.5, 1.25, 0.625, 0.3125, 0.156, 0.078, and 0 μM . Add 50 μL /well of serially diluted DMC from 5 μM to 0 nM into the 96-well plate or 20 μL /well into the 384-well plate.
- Add 50 μL /well MMP-7 substrate solution (refer to standard operation protocol step 3 for preparation) to the 96-well plate or 20 μL /well into the 384-well plate.

Note: The MMP-7 substrate solution should be added to the DMC reference standard to correct for the absorptive quenching by the FRET peptide. If multiple concentrations of substrate are used, this step must be performed for each concentration.

- *Optional:* If the stop solution (component E) was added into the enzymatic reaction before taking the end-point reading, the same volume of stop solution should be added to reference standard wells for better comparison.
- Plot DMC fluorescent reference standard as RFU (relative fluorescent unit) versus concentration.

Note: The final concentration of DMC reference standard is 2.5, 1.25, 0.625, 0.3125, 0.156, 0.078, 0.039, and 0 μM . This reference standard is used to calibrate the variation of different instruments and different batch of experiments. It is also an indicator of the amount of final product of the MMP-7 enzymatic reaction.

References

1. Woessner, JF. et al. *J. Biol. Chem.* **263**, 16918 (1988).
2. Woessner, JF. et al. *FASEB. J.* **5**, 2145 (1991).
3. Muller, D. et al. *Biochem J.* **253**, 187 (1988).
4. Stryer, L. et al. *Annu. Rev. Biochem.* **47**, 819 (1978).