ANASPEC

SensoLyte™ Anti-MOG (35-55) IgG Quantitative ELISA Kit (Mouse/Rat) *Colorimetric*

Revision number: 1.5   *Last updated: 27MAR2023*

Catalog #	AS-54465
Kit Size	One 96-well strip plate

This kit is optimized to detect mouse/rat anti-MOG (35-55) IgG. Wells are pre-coated with MOG (35-55) peptide and pre-blocked with BSA. The amount of anti-MOG (35-55) IgG in serum or cerebrospinal fluid is quantified using ELISA. Ample materials and reagents are provided to perform 96 assays.

- *Convenient Format*
    - Pre-coated and pre-blocked 96-well strip plate
    - Ready-to-use substrate solution and other assay components
    - 2-3 hours assay time at room temperature
- *Minimal Sample Size*
    - Requires only 0.5-1 µl of serum or cerebrospinal fluid to perform assay
- *High Sensitivity*
    - Detects as low as 100 pg anti-MOG (35-55) IgG
- *Broad Dynamic Range*
    - 8-500 ng antibody/ml serum (depending on colorimetric developing time)

Kit Components and Handling

Component	Description	Quantity
Component A	MOG (35-55) coated and BSA blocked 8-well strips	12 strips
Component B	Mouse anti-MOG (35-55) IgG standard	110 µl (5 µg/ml)
Component C	1X Sample Dilution Buffer	30 ml
Component D	10X Wash Buffer	50 ml
Component E	TMB color substrate solution	10 ml
Component F	Stop Solution	10 ml
Component G	Secondary antibody, Goat anti-Mouse IgG-HRP	30 µl
Component H	Secondary antibody, Goat anti-Rat IgG-HRP	30 µl
Component J	Rat anti-MOG (35-55) IgG standard	110 µl (5 µg/ml)

Other Materials Required (but not provided)

- Microplate reader: Capable of reading absorbance at 450 nm
- Rocking platform or shaker
- Strip ejector (to eject strips for future assay if not all strips are used in one experiment)
- Computer software: Capable of plotting Four Parameter Logistic Curve Fit (4-PL) (optional)

Shipment and Storage

- Kit is shipped on blue ice. Store all kit components at 2-8°C for up to 12 months.

For Research Use Only! Not for diagnostics.

Introduction

Myelin oligodendrocyte glycoprotein (MOG) is a member of the immunoglobulin superfamily and is expressed exclusively in the central nervous system.[1-3] MOG (35-55) is able to induce autoantibody production and relapsing-remitting neurological disease causing extensive plaque-like demyelination.[1-2] Autoantibody response to MOG (35-55) has been observed in multiple sclerosis (MS) patients and MOG (35-55)-induced experimental autoimmune encephalomyelitis (EAE) C57/BL6 mice and Lewis rats.[1-4]

The SensoLyte™ Anti-MOG (35-55) IgG Quantitative ELISA Kit (mouse/rat) provides a convenient and quantitative assay for anti-MOG (35-55) autoantibody in mouse and rat. This kit is useful to researchers for determining the amount of anti-MOG (35-55) antibody present, and can help provide information on the role it plays in the development and treatment of EAE,[4-5] an animal model for MS pathogenesis.

Protocol

Please Note:
      a) Allow kit components to warm up to room temperature before starting the assay
      b) Spin down all components with volume less than 100 µl before use
      c) Mix well 10X Washing Buffer before diluting to dissolve any precipitated salt
      d) More Sample Dilution Buffer can be made by adding 1% BSA into 1 X Wash Buffer

1.   ELISA assay:

1.1 Establish dilution range of serum samples: Serial dilutions of serum samples can start from 1:1k, 1:5k, 1:25k, 1:125k. Use 1X Sample Dilution Buffer (Component C) to dilute samples and standards (an example is shown in Table 1). Depending on the amount of antibody present, the dilution range can be further adjusted.

1.2 Arrange and label strips (Component A) based on the number of wells with standard and samples. An example is shown in Table 1. Although diluted standard and samples can be run as single points, duplicates are recommended.

Table 1. An example of four samples layout in duplicates using 6 eight-well strips.

	Standard [ng/ml]	Standard [ng/ml]	3	4	5	6
A	500	500	1:1K	1:1K	1:1K	1:1K
B	250	250	1:5K	1:5K	1:5K	1:5K
C	125	125	1:25K	1:25K	1:25K	1:25K
D	62.5	62.5	1:125K	1:125K	1:125K	1:125K
E	31.25	31.25	1:1K	1:1K	1:1K	1:1K
F	15.625	15.625	1:5K	1:5K	1:5K	1:5K
G	7.8125	7.8125	1:25K	1:25K	1:25K	1:25K
H	Blank	Blank	1:125K	1:125K	1:125K	1:125K

1.3   Depending on the animal model used, dilute mouse or rat anti-MOG (35-55) IgG standard
      (Component B or J) in 1X Sample Dilution Buffer (Component C) according to the Table 2.

Table 2. Serial dilution of anti-MOG (35-55) IgG standard.

Step	Concentration [ng/ml]	Anti-MOG IgG standard (Component B or J)	Sample Dilution Buffer (Component C)
1	500.00	100 µl from the stock	900 µl
2	250.00	500 µl from step 1	500 µl
3	125.00	500 µl from step 2	500 µl
4	62.5	500 µl from step 3	500 µl
5	31.25	500 µl from step 4	500 µl
6	15.625	500 µl from step 5	500 µl
7	7.812	500 µl from step 6	500 µl

1.4   Add 100 µl of the diluted standards into wells ($A_{1,2}$-$G_{1,2}$ for duplicate run). Add 100 µl of 1X
      Sample Dilution Buffer (Component C) as a blank into wells $H_{1,2}$.
1.5   Add diluted samples into appropriate wells (depends on the number of samples to be tested).
      After adding the standards and samples to the wells, cover the plate and incubate at room
      temperature for 60 min with gentle shaking.
1.6   Prepare 1X working wash buffer by diluting the 10X Wash Buffer (Component D) with DI H_2O.
1.7   Wash wells five times at 200 µl/well of 1X washing buffer. Pat dry.
1.8   Dilute goat anti-mouse IgG-HRP (Component G) secondary antibody (2nd Ab) with Sample
      Dilution Buffer (Component C) if quantifying mouse samples or/and goat anti-rat IgG-HRP
       (Component H) secondary antibody if quantifying rat samples. Secondary antibody working solution
      is 1:2,000 dilution. Add 100 µl of the diluted 2nd Ab into each well and incubate plate at room
      temperature for 45-60 min with gentle shaking.
1.9   Wash wells five times with 200 µl per well of 1X washing buffer. Pat dry. Clean the outside
      bottom of the wells with lens paper if necessary before the next step (this ensures accurate
      absorbance reading).
1.10 Add 100 µl of the TMB color substrate solution (Component E) into each well. Tap plate gently
      and incubate at room temperature until blue gradient is clearly observed across the wells
      (1-15 min depending on the secondary antibody dilution). It may be necessary to adjust color
      development time so that absorbance values fall within the detection range.
1.11 Add 50 µl of the Stop Solution (Component F) into each well and tap plate gently (blue color
      will turn to yellow). Measure absorbance (OD) at 450 nm using a microplate absorbance
      reader within 20 minutes after adding stop solution.

2.   Calculate the concentration of the samples.

2.1   Determine the average values (if replicates are used) for the standard and sample absorbance
      readings. Plot calibration curve using Four Parameter Logistic (4-PL) curve-fit. R^2 should be higher
      than 0.99. There should be at least 5 serially diluted standard concentrations in the calculation to
      ensure statistical significance.

2.2 Choose absorbance values for the samples that are within the range used in the standard curve, and calculate the concentration of anti-MOG (35-55) IgG using 4-PL curve-fit.

2.3 Example of calculation of anti-MOG (35-55) IgG concentrations:

Please note, new standard curve must be generated each time the assay is run.

Table 3. An example of the assay with 8 samples using mouse and rat standards.

	1	2	3	4	5	6	7	8	9	10	11	12
A	3.493	3.460	3.570	3.603	2.958	3.046	3.383	3.242	3.024	3.081	3.276	3.242
B	2.644	2.532	**2.089**	**2.197**	**1.678**	**1.698**	3.022	2.977	**2.056**	**2.088**	2.366	2.389
C	2.057	1.944	0.955	1.059	0.670	0.738	2.705	2.714	0.927	0.928	**1.179**	**1.177**
D	1.418	1.329	0.351	0.383	0.260	0.269	2.158	2.233	0.266	0.272	0.340	0.361
E	0.977	0.898	**1.568**	**1.664**	3.517	3.546	1.570	1.471	3.001	3.019	3.140	3.186
F	0.592	0.527	0.574	0.591	**2.108**	**2.184**	0.892	0.898	**1.762**	**1.822**	**1.817**	**1.849**
G	0.322	0.285	0.177	0.198	1.033	1.093	0.510	0.522	0.568	0.582	0.656	0.683
H	0.017	0.010	0.069	0.082	0.418	0.410	0.013	0.009	0.165	0.162	0.172	0.186

Note: Columns 1 and 2 are duplicate mouse anti-MOG(35-55) IgG standards 500, 250, 125, 62.5, 31.25, 15.625, 7.8, and 0 ng/ml (Row A ~ H). Mouse Samples: Sample-1, 3A-D and 4A-D; Sample-2, 5A-D and 6A-D; Sample-3, 3E-H and 4E-H; Sample-4, 5E-H and 6E-H (at 1:1k, 5k, 25k, and 125k dilution in duplicates). Columns 7 and 8 are duplicate rat anti-MOG(35-55) IgG standards 500, 250, 125, 62.5, 31.25, 15.625, 7.8, and 0 ng/ml (Row A ~ H). Rat Samples: Sample-1, 9A-D and 10A-D; Sample-2, 11A-D and 12A-D; Sample-3, 9E-H and 10E-H; Sample-4, 11E-H and 12E-H (at 1:1k, 5k, 25k, and 125k dilution in duplicates). Values in bold were used for calculations.

2.3.1 Four-parameter logistic curve-fit (4-PL) based on the average absorbance reading values:

Anti-Mouse MOG (35-55) IgG Standard Curve

Anti-Rat MOG (35-55) IgG Standard Curve
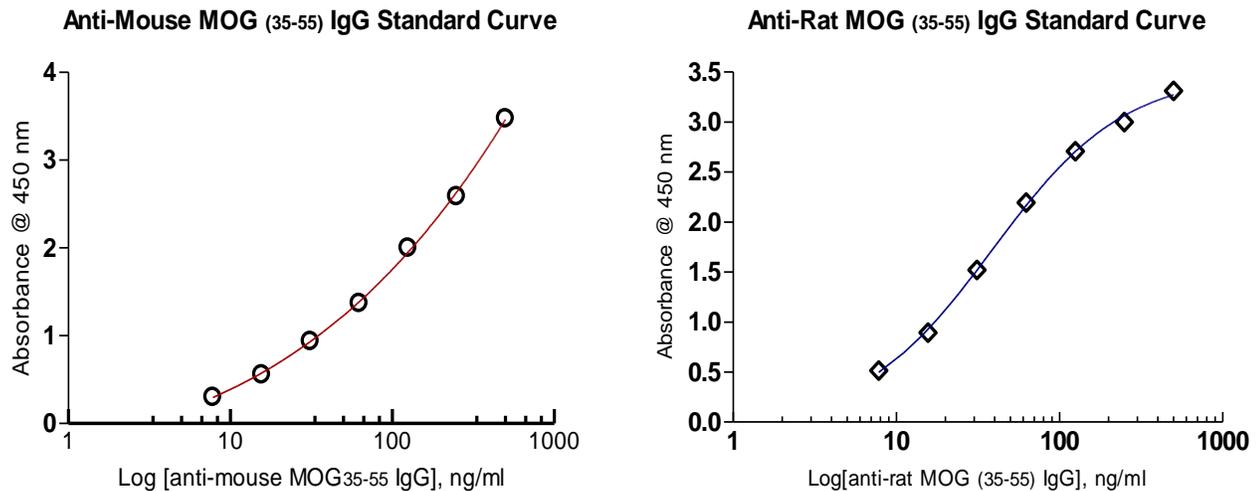
$Y=[(A-D)/(1+\{x/C\}^B)]+D$ $\underline{A}=-0.66$ $\underline{B}=0.39$ $\underline{C}=14842$ $\underline{D}=19.26$ $R^2=0.999$ $Y=[(A-D)/(1+\{x/C\}^B)]+D$ $\underline{A}=-0.03$ $\underline{B}=1.076$ $\underline{C}=38.88$ $\underline{D}=3.483$ $R^2=0.999$

2.3.2   From 4-PL curve-fit data table (not shown) generated by computer software the following anti-MOG (35-55) concentrations for mouse samples were obtained (based on the average absorbance readings):

	Absorbance @ 450nm, mean value	Calculated Concentration [ng/ml]	Dilution Factor	Actual Sample Concentration [mg/ml]
Sample1	2.143	155.52	1:5000	0.777
Sample2	1.688	93.04	1:5000	0.465
Sample3	1.616	85.81	1:1000	0.086
Sample4	2.146	155.7	1:5000	0.778

2.3.3   From 4-PL curve-fit data table (not shown) generated by computer software the following anti-MOG (35-55) concentrations for rat samples were obtained (based on the average absorbance readings):

	Absorbance @ 450nm, mean value	Calculated Concentration [ng/ml]	Dilution Factor	Actual Sample Concentration [mg/ml]
Sample1	2.072	56.92	1:5000	0.285
Sample2	1.178	21.27	1:25000	0.532
Sample3	1.792	42.88	1:5000	0.214
Sample4	1.833	43.86	1:5000	0.219

References:

1. Slavin, A. et al. *Autoimmunity* **28**, 109 (1998).
2. Ichikawa, M. et al. *Cellular Immunol.* **191**, 97 (1999).
3. Kennel De March, A. et al. *J. Neuroimmunol.* **135**, 117 (2003).
4. Robinson, W. et al. *Nat. Biotech.* **21**, 1033 (2003).
5. Mayo, S. et al. *Clin. Immunol.* **118**, 219 (2006).